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Protein A SuperSpin

Data and Instructions



hosts.

Protein A SuperSpin is supplied for fast small-scale purification of immunoglobulins from various species (e.g. serum, ascites and mammalian cell culture supernatants etc) at low cost. Protein A has been well proven a powerful affinity ligand that shows very high specificity to IgGs from mammalian

SuperSpin Tube

SuperSpin Tube

1.5 ml tube (not supplied)

Filter material

Each product consists of: 3 ml of Protein A SuperSpin Resin, 50 SuperSpin Tube, 50 pipetting tips.

Each Protein A SuperSpin tube accommodates 50 μ l of Protein A agarose particles. The affinity resin is specifically designed to provide a large accessible pore size with small particle size (i.e. 30 – 100 μ m). This means large antibody molecules can access the

affinity ligand rapidly with an excellent mass transfer rate. As a result, up to 0.5 milligram of antibody can be easily purified within 20 minutes at aninexpensive cost. No special expensive chromatography instrument or column is required.

Protein A SuperSpin is particularly useful for the following applications:

- If the quantity of antibody to be purified is small (up to 0.5 milligram)
- On-line detection of antibody expression levels
- High throughput screening of antibodies

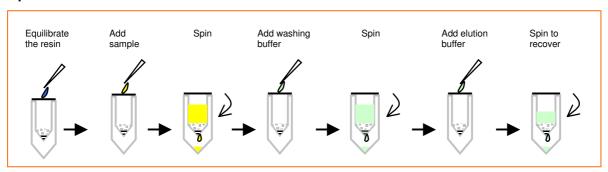
Key benefits:

- Rapid antibody capture and recovery
- High antibody binding capacity (up to 0.5 milligram)
- High throughput format (24 samples can be processed in parallel)
- Low cost
- Easy to use and no expensive equipment required

Product characteristics

Tube material	Polypropylene	
Resin	Protein A SuperSpin resin	
Packed volume	50 μl	
Protein binding capacity	Depends on the source of antibodies; up to 0.5 mg human IgG	
pH stability	3-9 in long term and $2-10$ in short term	
Chemical stability	The resin maintains the binding capability when exposed to 3M NaSCN,	
	6M GuHCl, 8 M urea and 70% ethanol for a few hours	
Storage condition	$4^{\circ}\text{C} - 30^{\circ}\text{C}$	

Operation Instructions



Note: standard 1.5 ml centrifuge tubes and a benchtop microcentrifuge are required in advance.

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- 1. Fully re-slurry the Protein A SuperSpin resin by gently shaking the resin bottle.
- 2. Using the yellow pipette tip supplied, pipette 100 µl of the slurry to each empty SuperSpin tube.
- 3. Place the SuperSpin tube in a standard 1.5 ml microcentrifuge tube (the lid of the 1.5 ml tube can be cut off). Spin at 1000 rpm for 30 seconds to settle the resin.
- 4. Take out the SuperSpin tube. Empty the 1.5 ml collection tube. Open the lid of the SuperSpin tube and load $200 \,\mu l$ of the binding buffer.
- 5. Close the lid of the SuperSpin tube. Spin in the collection tube at 6,500 rpm for 1 minute.
- 6. Place the SuperSpin tube into another fresh 1.5 ml tube. Add up to 200 μl of the protein sample and spin at 6,500 rpm for 1 minute. More of the sample can be added after each spin as long as it doesn't exceed the binding capacity (i.e. 0.5 mg). Make sure to fully empty the 1.5 ml tube after each spin. (Note: depending on the nature of individual sample, the spin speed and spin time may be vary to fully discharge the liquid.)
- 7. Place the SuperSpin tube into another fresh 1.5 ml tube. Add 200 µl of the washing buffer and spin at 6,500 rpm for 1 minute. The washing may be repeated one more time. Remember to empty the 1.5 ml tube after each spin.
- 8. Place the SuperSpin tube into another fresh 1.5 ml tube. Add 150 µl of the elution buffer (e.g. 0.1 M glycine buffer at pH 3.0). Spin at 6,500 rpm for 1 minute. The elution can be repeated one more time by adding another 100 µl of the elution buffer. Two elution fractions can be collected into the same tube or into two separate tubes. Most of the bound antibody will be eluted in the first fraction.

Ordering information

Product	Quantity	Code no.
Protein A SuperSpin	50 / pack	230102
Related products	Quantity	Code no.
SuperSpin Desaltor	50	210101

BioToolomics Ltd Unit 30A, Number 1 Industrial Estate Consett County Durham, DH8 6TJ United Kingdom

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